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Journal of Asian Natural Products Research

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713454007>

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To cite this Article Wu, Xian-Fu , Li, Yong , Lu, Hai-Ning , Yu, Shi-Shan , Ma, Shuang-Gang and Liu, Jing(2009) 'Prenylated C₆-C₃ compounds from the fruits of *Illicium simonsii*', Journal of Asian Natural Products Research, 11: 12, 1056 – 1061

To link to this Article: DOI: 10.1080/10286020903376194

URL: <http://dx.doi.org/10.1080/10286020903376194>

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Prenylated C₆–C₃ compounds from the fruits of *Illicium simonsii*

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(Received 11 September 2009; final version received 29 September 2009)

Two new prenylated C₆–C₃ compounds, 4-*epi*-illicinone E-12-shikimate (**1**) and 3-hydroxyillifunone B (**2**), together with five known prenylated C₆–C₃ compounds (**3**–**7**), were isolated from the fruits of *Illicium simonsii*. Their structures were elucidated on the basis of extensive spectroscopic methods, including 1D and 2D NMR, CD spectra, and ESI-MS analysis.

Keywords: *Illicium simonsii*; prenylated C₆–C₃ compounds; 4-*epi*-illicinone E-12-shikimate; 3-hydroxyillifunone B

1. Introduction

Prenylated C₆–C₃ compounds, also named phytoquinoids, were isolated frequently from *Illicium* plants in the previous reports [1–9]. *Illicium simonsii* belongs to the genus *Illicium* and is mainly distributed in Guizhou, Sichuan, and Yunnan Provinces of China, India, and Myanmar [10]. Since there are only a few reports concerning the chemical constituents of this plant [11,12], the fruits of *I. simonsii* were studied as part of an ongoing phytochemical investigation of the genus *Illicium*. As a result, seven prenylated C₆–C₃ compounds, including two new compounds (**1** and **2**), were isolated (Figure 1). In this paper, we report the isolation and structural elucidation of the two new compounds.

2. Results and discussion

Compound **1** was obtained as a colorless oil with the molecular formula C₂₂H₂₈O₉ established by positive HR-ESI-MS at *m/z* 437.1817. The IR spectrum showed

absorption bands attributable to hydroxyl groups (3404 cm⁻¹), one ester carbonyl (1708 cm⁻¹), and one α,β-conjugated carbonyl (1685 cm⁻¹). The ¹H and ¹³C NMR spectra of **1** (Table 1) revealed the presence of an allyl group [δ_H 2.96 (2H, d, *J* = 6.5 Hz, H₂-7), 5.82 (1H, m, H-8), 5.07 (1H, dd, *J* = 10.5, 1.5 Hz, H-9a), and 5.04 (1H, dd, *J* = 17.5, 1.5 Hz, H-9b); δ_C 33.7 (C-7), 135.8 (C-8), and 117.4 (C-9)], an α,β-conjugated carbonyl group [δ_H 6.63 (1H, s, H-3); δ_C 194.5 (C-1), 138.9 (C-2), and 139.0 (C-3)], two isolated methylene groups [δ_H 3.09 (1H, d, *J* = 16.5 Hz, H-6a), 2.92 (1H, d, *J* = 16.5 Hz, H-6b), 2.51 (1H, dd, *J* = 14.0, 5.0 Hz, H-10a), and 2.23 (1H, dd, *J* = 14.0, 11.0 Hz, H-10b); δ_C 46.9 (C-6) and 38.4 (C-10)], two dimethyl groups linked to a quaternary carbon [δ_H 1.47 (3H, s, H₃-13) and 1.51 (3H, s, H₃-14); δ_C 22.0 (C-13) and 22.5 (C-14)], and a methylenedioxy group [δ_H 4.92 (1H, s, H-15a) and 5.16 (1H, s, H-15b); δ_C 95.3 (C-15)]. The above NMR spectral

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Table 1. ^1H (500 MHz) and ^{13}C (125 MHz) NMR spectral data for compounds **1** and **2** in acetone- d_6 .

No.	1		2	
	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}
1		194.5		199.2
2		138.9	3.56 m	44.2
3	6.63 s	139.0	4.15 d (3.0)	72.4
4		86.0		78.5
5		111.9		180.4
6	3.09 d (16.5) 2.92 d (16.5)	46.9	5.21 s	96.4
7	2.96 d (6.5)	33.7	2.44 dd (7.5, 14.0) 2.32 ddd (5.0, 7.0, 14.0)	41.8
8	5.82 m	135.8	5.87 m	133.9
9	5.07 dd (10.5, 1.5) 5.04 dd (17.5, 1.5)	117.4	5.10 dd (10.0, 1.5) 5.04 dd (17.0, 1.5)	118.2
10	2.51 dd (14.0, 5.0) 2.23 dd (14.0, 11.0)	38.4	2.36 dd (5.0, 11.5) 2.10 dd (11.0, 11.5)	26.3
11	4.52 dd (11.0, 5.0)	85.7	4.52 dd (5.0, 11.0)	93.3
12		82.1		70.4
13	1.47 s	22.0	1.27 s	26.5
14	1.51 s	22.5	1.17 s	25.6
15	4.92 s 5.16 s	95.3		
1'		131.0		
2'	6.61 br s	138.4		
3'	4.34 br s	66.7		
4'	3.66 dd (4.0, 7.0)	72.3		
5'	3.97 m	68.0		
6'	2.07 dd (5.0, 19.0) 2.55 dd (5.0, 19.0)	31.4		
7'		166.2		

$J = 7.5, 14.0$ Hz, H-7b), 5.87 (1H, m, H-8), 5.04 (1H, dd, $J = 10.0, 1.5$ Hz, H-9a), and 5.10 (1H, dd, $J = 17.0, 1.5$ Hz, H-9b); δ_{C} 41.8 (C-7), 133.9 (C-8), and 118.2 (C-9)], a dimethyl carbinol group [δ_{H} 1.27 (3H, s, H₃-13) and 1.17 (3H, s, H₃-14); δ_{C} 70.4 (C-12), 26.5 (C-13), and 25.6 (C-14)], and an α,β -conjugated carbonyl

[δ_{H} 5.21 (1H, s, H-6); δ_{C} 199.2 (C-1), 180.4 (C-5), and 96.4 (C-6)]. The NMR spectral data of **2** were similar to those of illifunone B [1], except that the signal at δ_{C} 34.9 (C-3) in illifunone B was shifted downfield to δ_{C} 72.4 (C-3) in **2**, which disclosed that one proton at C-3 in illifunone B was replaced by a hydroxyl

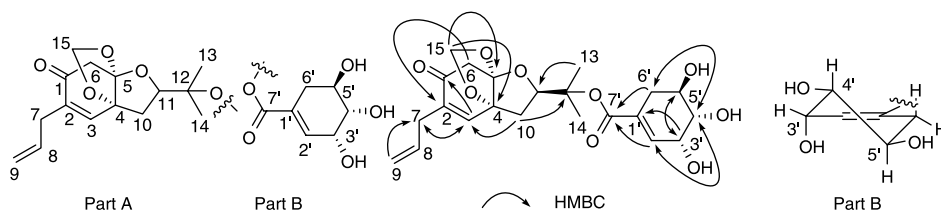


Figure 2. Structures of parts A and B, key HMBC correlations of **1**, and Haworth projection of part B.

group in **2** in combination with their molecular formula.

The relative configuration of **2** was established by the chemical shift of H-11 and the NOE experiment (Figure 3). A *syn*-relationship between the hydroxyl group at C-4 and the dimethyl carbinol group at C-11 was elucidated from the chemical shift of H-11 (δ_{H} 4.52) appearing at higher field than δ_{H} 4.6 [7]. Moreover, an *anti*-relationship between the allyl group at C-2 and the hydroxyl group at C-3 was confirmed by the observation of NOE for H-3 upon irradiation of H-7b (δ_{H} 2.44). In addition, a *syn*-relationship between the allyl group at C-2 and the dimethyl carbinol group at C-11 was established for the NOEs between H-2 and H-11. The absolute configuration of C-2 was determined to be *R* on the basis of the positive Cotton effect at 310 nm [7]. Therefore, the absolute configuration for compound **2** was deduced as 2*R*, 3*R*, 4*S*, 11*R*. Thus, the structure of **2** was determined as shown in Figure 1.

The structures of the other five known prenylated C₆-C₃ compounds, 6-allyl-6-(3-methyl-2-butenyl)-3,4-methylenedioxy-cyclohexa-2,4-dienone (**3**) [3], illicinone E (**4**) [16], illifunone C (**5**) [1,7], illifunone D (**6**) [1,7], and 2,3-dehydroillifunone C (**7**) [7], were elucidated by comparison of their spectral data with those reported.

3. Experimental

3.1 General experimental procedures

Optical rotations were determined on a Perkin-Elmer 241 automatic digital polarimeter. A CD spectrum was obtained from a JOUAN Mark II spectropolarimeter. IR spectra were recorded on a Nicolet 5700 FT-IR spectrometer. 1D and 2D NMR spectra were recorded on a Varian INOVA-500 spectrometer with TMS as an internal standard. ESI-MS were measured on an Agilent 1100 series LC/MSD trap mass spectrometer. HR-ESI-MS spectra were recorded on an Autospec-Ultima ETOF Spec mass spectrometer. Preparative HPLC was performed on a Shimadzu LC-6AD instrument with an SPD-10A detector. Silica gel GF₂₅₄ for TLC was obtained from Qingdao Marine Chemical Company, Qingdao, China. ODS (50 μ) and Sephadex LH-20 were purchased from Fuji Silysica Chemical Ltd (Greenville, NC, USA).

3.2 Plant material

The fruits of *I. simonsii* were collected from Yunnan Province of China, which was identified by Prof. Lin Ma of the Institute of Materia Medica, Chinese Academy of Medical Sciences and Peking Union Medica, where a voucher specimen (No. 90204) has been deposited.

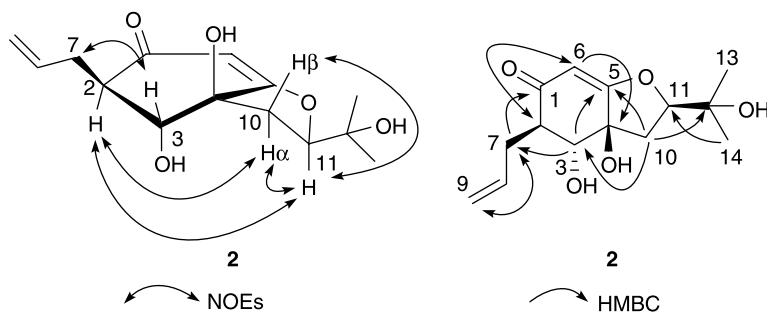


Figure 3. Key NOE and HMBC correlations of **2**.

3.3 Extraction and isolation

The dried powder of the fruits of *I. simonsii* (1.1 kg) was extracted with 95% EtOH (15 liters \times 3) and concentrated *in vacuo* to give the crude extract (158 g), which was absorbed by kieselguhr, and then successively extracted with petroleum ether, CHCl_3 , EtOAc, and MeOH. The CHCl_3 extract (25 g) was subjected to the ODS column eluted with MeOH– H_2O (from 65:35 to 85:15) to give five fractions (A–E). Fraction B (2.6 g) was subjected to Sephadex LH-20 (CH_2Cl_2 –MeOH, 1:1) and then further purified on preparative HPLC (MeOH– H_2O , 84:16) to yield compounds **1** (13.1 mg), **2** (9.1 mg), and **7** (16.1 mg). Fraction E (3.3 g) was chromatographed on the ODS column, eluted with a gradient system of MeOH– H_2O (from 60:40 to 90:10), and further separated by preparative HPLC (MeOH– H_2O , 88:12) to afford compounds **3** (4.1 mg), **4** (9.6 mg), **5** (12.4 mg), and **6** (6.1 mg).

3.3.1 Compound 1

Colorless oil; $[\alpha]_{\text{D}}^{20}$ –38.1 ($c = 0.08$, MeOH); UV (MeOH) λ_{max} : 225 nm; CD (MeOH) $\Delta\epsilon_{200 \text{ nm}}$ 18.2, $\Delta\epsilon_{224 \text{ nm}}$ 0, $\Delta\epsilon_{320 \text{ nm}}$ –1.66, $\Delta\epsilon_{325 \text{ nm}}$ 0; IR (KBr) ν_{max} : 3404, 2981, 2907, 1708, 1685, 1613, 1369, 1235, 1137, 1095, 1072 cm^{-1} ; ^1H and ^{13}C NMR spectral data, see Table 1; positive HR-ESI-MS m/z : 437.1817 $[\text{M}+\text{H}]^+$ (calcd for $\text{C}_{22}\text{H}_{29}\text{O}_9$, 437.1806).

3.3.2 Compound 2

Colorless oil; $[\alpha]_{\text{D}}^{20}$ –68.3 ($c = 0.05$, MeOH); UV (MeOH) λ_{max} : 265 nm; CD (MeOH) $\Delta\epsilon_{201 \text{ nm}}$ 0, $\Delta\epsilon_{256 \text{ nm}}$ –11.8, $\Delta\epsilon_{296 \text{ nm}}$ 0, $\Delta\epsilon_{310 \text{ nm}}$ 1.04, $\Delta\epsilon_{348 \text{ nm}}$ 0; IR (KBr) ν_{max} : 3429, 2988, 2911, 1657, 1624, 1389, 1376, 1190, 1079 cm^{-1} ; ^1H and ^{13}C NMR spectral data, see Table 1; positive HR-ESI-MS m/z : 291.1209 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{14}\text{H}_{20}\text{O}_5\text{Na}$, 291.1203).

3.4 Alkaline hydrolysis of 1

Compound **1** (6.2 mg) was hydrolyzed with 0.5 M NaOH (3 ml) and MeOH (0.5 ml) for 1 h at room temperature. After adjusting the pH to 5.0 with 0.5 M HCl, the reaction mixture was extracted with EtOAc, which was separated by HPLC (CH_3CN – H_2O , 18:92) to afford (–)-shikimic acid (1.2 mg).

3.4.1 (–)-Shikimic acid

Colorless powder; $[\alpha]_{\text{D}}^{20}$ –138.1 ($c = 0.05$, MeOH); ^1H NMR (500 MHz, acetone- d_6): δ_{H} 6.71 (1H, m, H-2), 4.40 (1H, t, $J = 4.0$ Hz, H-3), 3.68 (1H, dd, $J = 7.0, 4.0$ Hz, H-4), 3.99 (1H, m, H-5), 2.68 (1H, dd, $J = 19.0, 5.0$ Hz, H-6a), 2.11 (1H, dd, $J = 19.0, 5.0$ Hz, H-6b); ^{13}C NMR (125 MHz, acetone- d_6): δ_{C} 138.1 (C-1), 133.7 (C-2), 71.6 (C-3), 68.1 (C-4), 67.2 (C-5), 31.3 (C-6), 170.1 (C-7).

Acknowledgements

The project was supported by the National Science Fund for Distinguished Young Scholars (No. 30625040) and the National Key Basic R&D (973) Project (No. 2004CB518906). We thank the Department of Instrumental Analysis, Institute of Materia Medica, Chinese Academy of Medical Sciences and Peking Union Medical College for measuring the IR, NMR, MS, and CD spectra.

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